Sesquiterpenes and Lignans from Tephrosia vogelii

by Huan-Huan Wei^a)^b), Han-Hong Xu^c), Hai-Hui Xie^a), Liang-Xiong Xu^a), and Xiao-Yi Wei^{*a})

 ^a) CAS Key Laboratory for Preservation and Sustainable Utilization of Plant Resources, South China Botanical Garden, Chinese Academy of Science, Guangzhou 510650, P. R. China (phone: +86-20-37252538; fax: +86-20-37252537; e-mail: wxy@scbg.ac.cn)
^b) Key Laboratory of Molecular Medicine, Ministry of Education, Shanghai 200032, P. R. China
^c) Key Laboratory of Natural Pesticides and Chemical Biology, Ministry of Education, South China

Agriculture University, Wushan, Guangzhou 510640, P. R. China

A new guaiane sesquiterpene, $(1\beta,6\alpha,10\alpha)$ -guai-4(15)-ene-6,7,10-triol (1), and a new lignan, (+)lariciresinol 9'-stearate (2), were isolated from the aerial parts of a leguminosae plant, *Tephrosia vogelii*, together with three known compounds, teclenone B, $(1\beta,7R^*)$ -opposit-4(15)-ene-1,7-diol, and pinoresinol. Their structures were elucidated on the basis of spectral data.

Introduction. – *Tephrosia vogelii* HOOK f., a perennial leguminous shrub native to tropical Africa, is known as a source of the rotenoids including rotenone and deguelin [1][2]. It was introduced into agriculture in the southern area of Guangdong Province, P. R. China, in 1980s, where its aerial parts have been used for the commercial production of rotenoids [3]. The general process for the preparation of rotenoids from this plant yields, besides rotenoids, a large amount of nonrotenoid fractions. Currently, the rotenoids are applied to pesticides, while the nonrotenoid fractions are discarded as undesirable substances, which results not only in an adverse impact on the environment but also in the waste of a resource. Therefore, it is important to investigate the nonrotenoid constituents of *T. vogelii* so that they can be utilized as valuable compounds rather than just be discarded as waste. However, the nonrotenoid constituents of this plant are almost unknown. Herein, we report the isolation and structure elucidation of a new sesquiterpene, $(1\beta, 6\alpha, 10\alpha)$ -guai-4(15)-ene-6,7,10-triol¹) (1), and of a new lignan, (+)-lariciresinol 9'-stearate¹) (2), from this plant cultivated in South China.



1) Trivial atom numbering; for systematic names, see *Exper. Part.*

^{© 2009} Verlag Helvetica Chimica Acta AG, Zürich

Results and Discussion. – The EtOH extract of the aerial parts of *T. vogelii* was partitioned with petroleum ether, CHCl₃, AcOEt, and BuOH. The CHCl₃-soluble portion was separated by a combination of silica gel, *ODS*, and *Sephadex LH-20* column chromatography and semi-prep. HPLC to afford two new compounds, **1** and **2**, together with two known sesquiterpenes, teclenone B [4], $(1\beta,7R^*)$ -opposit-4(15)-ene-1,7-diol [5], and a known lignan, pinoresinol [6][7]. The structures of the known compounds were determined by interpretation of their spectroscopic data as well as by comparison with reported data.

Compound 1 was obtained as a colorless gum. Its molecular formula was determined to be C₁₅H₂₆O₃ by the combined analysis of its HR-ESI-MS, ¹³C-NMR, and DEPT data. The ¹H-NMR spectrum (see Exper. Part) exhibited signals for 23 nonexchangeable H-atoms, including two d at $\delta(H) 0.86 (J = 7.0 \text{ Hz}, \text{Me}(12))$ and 0.92 (J = 7.0 Hz, Me(13)) for two secondary Me groups, a s at $\delta(H)$ 1.10 (Me(14)) for a tertiary Me group, a d at $\delta(H)$ 3.25 (J = 10.0, H–C(6)) for an O–CH group, and two broad s at $\delta(H)$ 4.78 (H_a-C(15)) and 5.04 (H_b-C(15)) for an exocyclic olefinic CH₂ group. The ¹³C-NMR and DEPT spectra indicated the presence of three Me groups $(\delta(C) \ 16.5 \ (C(13)), \ 17.8 \ (C(12)), \ and \ 25.6 \ (C(14))), \ five \ CH_2 \ groups \ including \ one$ olefinic (δ (C) 109.0 (C(15))), four CH groups including one oxygenated (δ (C) 74.4 (C(6))), and three quaternary C-atoms, of which two were oxygenated (δ (C) 75.0 (C(7)) and 73.2 (C(10)) and one was obtained ($\delta(C)$ 154.3 (C(4))). The ¹H,¹H-COSY in combination with the HMQC spectrum revealed the partial structures shown by the bold lines in Fig. 1. In the HMBC spectrum (Fig. 1), correlations observed from H-C(1), H-C(5), $CH_2(8)$, $CH_2(9)$, and Me(14) to C(10), and from H-C(5), H-C(6), $CH_2(8)$, $CH_2(9)$, H-C(11), Me(12), and Me(13) to C(7) indicated the connectivity of C(10) to C(1) and C(9), and of C(7) to C(6) and C(8), forming a sevenmembered ring (B) with a Me group (C(14)) and an OH group attached at C(10) and an ⁱPr group and an OH group attached at C(7). HMBC Cross-peaks were also observed between the olefinic CH₂ H-atoms (H_a-C(15) and H_b-C(15)) and C(4), C(3), and C(5), showing the connection of C(4) to C(3) and C(5), forming a fivemembered ring (A) with a CH₂ (C(15)) attached at C(4) via a C=C bond. Thus, the planar structure of 1 was derived as guai-4(15)-ene-6,7,10-triol.



Fig. 1. ¹H,¹H-COSY (bold line) and key HMBC (arrow) data of 1 and 2

The relative configuration of **1** was determined from the NOE interactions in the NOESY plot (*Fig. 2*). The presence of the mutual NOE correlations H-C(6)/H-C(1),

 $H-C(6)/H_{\beta}-C(8), H-C(1)/H_{\beta}-C(8), Me(12)/H-C(6), Me(13)/H-C(6), Me(12)/$ $CH_{2}(8)$, Me(13)/CH₂(8), H-C(5)/Me(14), and Me(14)/H_a-C(9) and the absence of the correlations $Me(14)/H_{\beta}-C(9)$, Me(12)/H-C(1), $Me(12)/CH_2(9)$, Me(13)/H-C(1), $Me(13)/CH_2(9)$, and Me(14)/H-C(1) as well as the proton coupling constant, J(5,6) = 10.0 Hz, in the ¹H-NMR spectrum indicated that the fusion between rings A and B was *trans*, and the ring B was in a chair-like conformation with C(1), C(5), C(7), and C(8) lying in the same plane, while C(9) and C(10) were oriented below and C(6) above this plane (Fig. 2). These correlations further indicated that OH-C(6) was α -oriented and in equatorial position, and OH-C(7) and Me-C(10) were also α -oriented but in axial positions. Therefore, the structure of **1** was established as $(1\beta, 6\alpha, 10\alpha)$ -guai-4(15)-ene-6,7,10-triol. When the modified *Mosher* method [8] was applied to establish the absolute configuration, **1** failed to react with (aR)- or (aS)-amethoxy- α -(trifluoromethyl)benzeneacetic acid (MTPA) in the presence of 1-ethyl-3-[3-(dimethylamino)propyl]carbodiimide hydrochloride (EDC·HCl) and catalytic N,N-dimethylpyridin-4-amine (DMAP) [9], presumably due to the hindrance of the vicinal ⁱPr group at C(7). This was in accordance with the attributed relative configuration of **1** as shown in *Fig. 2*.



Fig. 2. Relative configuration and significant NOESY interactions of 1

Compound **2** was obtained as a pale yellow powder. Its molecular formula was established as $C_{38}H_{58}O_7$ from the HR-ESI-MS ion peak at m/z 649.4080 ($[M + Na]^+$). In the ¹H-NMR spectrum (see *Exper. Part*), the presence of a *t* at $\delta(H) 0.83$ (J = 6.8 Hz, Me(18'')) for a Me group, a *t* at $\delta(H) 2.19$ (J = 7.2 Hz, CH₂(2'')) assignable to a CH₂ group vicinal to a C=O group, and a number of *ms* with great intensity overlapping in the region $\delta(H) 1.10 - 1.30$ indicated that **2** had a fatty acid moiety in the molecule. This was supported by the presence of C-atom signals at $\delta(C) 172.8$ for an ester C=O group and $\delta(C) 13.9$ for a Me group, and partially overlapping peaks from $\delta(C) 22.1$ to 33.5 for a number of CH₂ in the ¹³C-NMR and DEPT spectra. After having attributed the signals due to the fatty acid moiety, the remaining NMR signals were very similar to those of (+)-lariciresinol (=(2S,3R,4R)-tetrahydro-2-(4-hydroxy-3-methoxyphenyl)-4-[(4-hy-1) and the state of t

droxy-3-methoxyphenyl)methyl]furan-3-methanol) [10] [11], except that the signals of CH₂(9') (δ (H) 4.25 and 4.06 (each *dd*, *J* = 11.2, 7.2 Hz, 1 H) and C(9') (δ (C) 62.0) of **2** were downfield-shifted. The relative molecular mass of **2**, 626, was 266 units more than that of lariciresinol, indicating that the fatty acid moiety was a stearoyl group. The presence of correlations between CH₂(9') and C(1'') (δ (C) 172.8) in the HMBC plot (*Fig. 1*) indicated that the stearoyloxy group was linked to C(9'). The configuration of **2** should be the same as that of (+)-lariciresinol, as deduced from its positive optical rotation and negative *Cotton* effects at 232 and 289 nm in the CD spectrum [12]. Thus, the structure of **2** was determined as (+)-lariciresinol 9'-stearate.

This work was supported by a project (2005B30101014) from the *Department of Science and Technology of Guangdong Province*, China, and a grant from the Key Laboratory of Natural Pesticides and Chemical Biology, Ministry of Education, South China Agriculture University, Guangzhou, P. R. China.

Experimental Part

General. Column chromatography (CC): silica gel 60 (SiO₂, 100–200 mesh; Qingdao Marine Chemical Inc., Qingdao, P. R. China), Develosil ODS (10 µm; Nomura Chemical Co. Ltd., Japan), and Sephadex LH-20 (Pharmacia Biotech AB, Uppsala, Sweden). Prep. HPLC: Shimazu-LC-6A pump, Shimazu-RID-10A refractive-index detector, and XTerra-prep-MS- C_{18} column (10 µm, 19 × 300 mm). Optical rotations: Perkin-Elmer 341 polarimeter. CD Spectrum: Jasco-810 spectropolarimeter; λ ([θ]) in nm. UV Spectra: Perkin-Elmer Lambda-25 UV/VIS spectrophotometer; λ_{max} (log ε) in nm. NMR Spectra: Bruker-DRX-400 instrument; at 400 (¹H) or 100 MHz (¹³C); in CDCl₃ or (D₆)DMSO; δ in ppm rel. to Me₄Si, J in Hz. ESI-MS: MDS-SCIEX-API-2000 LC/MS/MS instrument. HR-ESI-MS: API-QSTAR-TOF mass spectrometer.

Plant Material. The aerial parts of *T. vogelii* were collected at the garden for pesticidal plants, South China Agricultural University, Guangzhou, China, in June, 2006. The voucher specimens were deposited with the Key Laboratory of Natural Pesticides and Chemical Biology, Ministry of Education, South China Agriculture University, Guangzhou, China.

Extraction and Isolation. The powdered dry stems and leaves of *T. vogelii* (2.5 kg) were extracted with 90% EtOH (4×) at r.t. After evaporation of the solvent, the extract was suspended in H₂O and then partitioned sequentially with petroleum ether, CHCl₃, AcOEt, and BuOH. The CHCl₃-soluble part (94.6 g) was subjected to CC (SiO₂, petroleum ether/acetone $19:1 \rightarrow 4:1$): *Frs. I–VIII. Fr. IV* was further subjected to CC (SiO₂, CHCl₃/MeOH $9:1 \rightarrow 7:3$): *Frs. IV-1 – IV-7. Fr. IV-2* was separated by CC (*ODS*, 70% MeOH/H₂O) followed by purification by prep. HPLC (60% MeOH/H₂O) to afford **1** (35 mg), teclenone B (52 mg), and (1β , *TR**)-opposit-4(15)-ene-1,7-diol (17 mg). By the same method, *Fr. IV-5* afforded **2** (21 mg), and *Fr. IV-7* gave pinoresinol (66 mg).

 $\begin{array}{l} (l\beta, 6a, 10a) - Guai + (15) - ene - 6, 7, 10 - triol (= rel-(3aR, 4R, 5R, 8S, 8aS) - Decahydro-8-methyl-3-methyl$ ene -5-(1-methylethyl)azulene -4, 5, 8-triol;**1** $). Colorless gum. [a]_{20}^{20} = + 20.0 (c = 0.25, MeOH). ¹H-NMR (400 MHz, CDCl₃): 5.04 (br. s, H_b - C(15)); 4.78 (br. s, H_a - C(15)); 3.25 (d, J = 10.0, H - C(6)); 2.31 (br. dd, J = 14.0, 6.4, H_a - C(3)); 2.25 (m, H_β - C(3)); 2.23 (dd, J = 10.0, 8.8, H - C(5)); 2.13 (m, H - C(11)); 2.10 (ddd, J = 14.0, 10.8, 2.0, H_a - C(9)); 1.93 (m, H - C(1)); 1.86 (m, H_β - C(2)); 1.67 (br. dd, J = 15.2, 11.2, H_β - C(8)); 1.57 (ddd, J = 15.2, 6.8, 6.8, H_a - C(8)); 1.43 - 1.50 (m, H_a - C(2), H_β - C(9)); 1.10 (s, Me(14)); 0.92 (d, J = 7.0, Me(13)); 0.86 (d, J = 7.0, Me(12)). ¹³C-NMR (100 MHz, CDCl₃): 154.3 (C(4)); 109.0 (C(15)); 75.0 (C(7)); 74.4 (C(6)); 73.2 (C(10)); 51.0 (C(1)); 46.4 (C(5)); 35.8 (C(9)); 34.9 (C(3)); 34.8 (C(11)); 27.3 (C(2)); 25.6 (C(14)); 21.4 (C(8)); 17.8 (C(12)); 16.5 (C(13)). ESI-MS (pos.): 531 ([2M + Na]⁺), 293 ([M + K]⁺), 277 ([M + Na]⁺). HR-ESI-MS: 277.1773 ([M + Na]⁺, C₁₅H₂₆NaO₃⁺; calc. 277.1780).$

(+)-Lariciresinol 9'-Stearate (= Octadecanoic Acid {(2\$,3\$,4\$,R})-Tetrahydro-2-(4-hydroxy-3-methoxy-phenyl)-4-[(4-hydroxy-3-methoxyphenyl)methyl]furan-3-yl]methyl Ester; 2). Pale yellow amorphous

powder. $[a]_{D}^{20} = +8.1$ (c = 1.0, MeOH). CD ($c = 1 \cdot 10^{-4}$ M, MeOH): 232 (-10362), 289 (-1980). UV (MeOH): 220 (4.37), 277 (4.00). ¹H-NMR (400 MHz, (D₆)DMSO): 6.82 (d, J = 1.6, H-C(2')); 6.74 (d, J = 1.6, H-C(2)); 6.70 (d, J = 8.0, H-C(5')); 6.67 (dd, J = 8.0, 1.6, H-C(6')); 6.66 (d, J = 8.0, H-C(5')); 6.55 (dd, J = 8.0, 1.6, H-C(6')); 6.66 (d, J = 8.0, H-C(5')); 6.55 (dd, J = 8.0, 1.6, H-C(6')); 3.89 (dd, J = 8.2, 6.6, 1 H-C(9')); 3.73 (s, MeO-C(3), MeO-C(3')); 3.56 (dd, J = 8.2, 6.6, 1 H-C(9)); 2.72 (dd, J = 13.4, 4.6, H_a-C(7)); 2.65 (m, H-C(8)); 2.47 (m, H_b-C(7)); 2.41 (m, H-C(8')); 2.19 (t, J = 7.2, CH₂(2'')); 1.45 (m, CH₂(3'')); 1.10-1.30 (m, 28 H, CH₂(4'') to CH₂(17'')); 0.83 (t, J = 6.8, Me(18'')). ¹³C-NMR (100 MHz, (D₆)DMSO): 172.8 (C(1'')); 147.4 (C(3), C(3')); 145.8 (C(4')); 144.7 (C(4)); 133.5 (C(1')); 131.1 (C(1)); 120.5 (C(6)); 118.5 (C(6')); 115.3 (C(5)); 115.0 (C(5')); 112.7 (C(2)); 110.1 (C(2')); 82.3 (C(7')); 71.8 (C(9)); 62.0 (C(9')); 55.5 (MeO -C(3), MeO -C(3')); 84.6 (C(8')); 42.0 (C(8)); 33.5 (C(2'')); 31.3 (C(16'')); 32.4 (C(7)); 29.0 - 28.5 (C(4'') to C(15'')); 24.4 (C(3'')); 22.1 (C(17'')); 13.9 (C(18'')). ESI-MS (pos.): 1275 ([2M + Na]⁺), 649 ([M + Na]⁺). HR-ESI-MS: 649.4080 ([M + Na]⁺, C₃₈HaO⁺; calc. 649.4080).

REFERENCES

- [1] H. Matovu, D. Olila, Int. J. Trop. Med. 2007, 2, 83.
- [2] J. W. Hagemann, M. B. Pearl, J. J. Higgins, N. E. Delfel, F. R. Earle, J. Agric. Food Chem. 1972, 20, 906
- [3] J. G. Huang, L. J. Zhou, H. H. Xu, Y. X. An, Plant Protection 2006, 32, 18.
- [4] A. J. Al-Rehaily, M. S. Ahmad, J. S. Mossa, I. Muhammad, J. Nat. Prod. 2002, 65, 1374.
- [5] T. Iijima, Y. Yaoita, M. Kikuchi, Chem. Pharm. Bull. 2003, 51, 545.
- [6] A. C. Casabuono, A. B. Pomillo, Phytochemistry 1994, 35, 479.
- [7] M. He, J. H. Zhang, C. Q. Hu, Acta Pharm. Sin. 2001, 36, 278.
- [8] I. Ohtani, T. Kusumi, Y. Kashman, H. Kakisawa, J. Am. Chem. Soc. 1991, 113, 4092.
- [9] N. Feng, W. Ye, P. Wu, Y. Huang, H. Xie, X. Wei, J. Antibiot. 2007, 60, 179.
- [10] C.-Y. Duh, C. H. Phoebe Jr., J. M. Pezzuto, A. D. Kinghorn, N. R. Farnsworth, J. Nat. Prod. 1986, 49, 706.
- [11] F. Abe, T. Yamauchi, Phytochemistry 1989, 28, 1737.
- [12] K. Machida, M. Takano, R. Kakuda, Y. Yaoita, M. Kikuchi, Chem. Pharm. Bull. 2002, 50, 669.

Received July 15, 2008